



Potassium 2,3,4-trihydroxy-2-methylbutanoate, a leaf-closing substance of *Leucaena leucocephalam*

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Received 29 January 2001; revised 26 February 2001; accepted 2 March 2001

Abstract—Potassium 2,3,4-trihydroxy-2-methylbutanoate (**1**) was identified as a leaf-closing substance of a nyctinastic plant, *Leucaena leucocephalam*. Compound **1** was quite effective for the leaf-closing of *L. leucocephalam* at 1×10^{-6} M. © 2001 Elsevier Science Ltd. All rights reserved.

Most legumes close their leaves in the evening, as if to sleep, and open them in the morning.¹ This rhythmic movement of the leaves is called nyctinasty, which has been known to be controlled by an internal biological clock.² Recently, we have identified several bioactive substances that regulate this leaf-movement, and revealed that nyctinastic movement of the plants is controlled by the interaction between leaf-closing and -opening substances, which are different among the plants.³ Moreover, we have demonstrated the importance of leaf-movement for the survival of legumes. In our previous work with artificial leaf-opening substance the leaf of *Cassia mimosoides* L. which was kept open for a week was observed to wither and die.⁴ This result showed that the nyctinastic leaf-movement is essential for the survival of legumes. From these findings, we envisioned that plant-specific leaf-movement factors could be useful as a herbicide. When we use such leaf-movement factor as a herbicide, it would be effective only for the legume from which it was isolated, and have no effect on other vicinal plants.

Nyctinasty is also observed in *Leucaena leucocephalam* (gin-nemu in Japanese), a leguminous tropical plant. *L. leucocephalam* is known for its rapid growth and secretion of allelochemicals to inhibit the growth of other plants around them, which leads to a grove of that plant and elimination of vicinal plants.⁵ Thus, the

disruption of an ecosystem by *L. leucocephalam* is a serious problem. We have tried to develop a herbicide, which is specifically effective for *L. leucocephalam*, by using the leaf-movement factor of this plant. This new type of highly ecological herbicide will exterminate only *L. leucocephalam* and, on the other hand, have no effect on other plants. Thus, we tried to identify the leaf-movement factors of *L. leucocephalam*. In this paper, we describe the isolation, chemical structure, and biological activity of the leaf-closing substance of *L. leucocephalam*.

Isolation of the leaf-closing substance was carried out based on a bioassay using a leaf of *L. leucocephalam*. The bioactive fraction closed a leaf within a few hours after the addition of a solution containing the bioactive substance.

The fresh whole plant of *L. leucocephalam* (13.0 kg) which was collected in Okinawa, Japan, was extracted with MeOH (72 L) for 1 week and concentrated in vacuo. The concentrated extract was partitioned with *n*-hexane, ethyl acetate, then with *n*-butanol. The bioactive aqueous layer was carefully separated by Amberlite XAD-7 column chromatography eluted with MeOH–H₂O (0:100, 10:90, 30:70, 50:50, and 100:0), and the H₂O fraction showed weak leaf-opening activity. This H₂O fraction was further purified with column chromatography using TSK G3000S gel with 30% EtOH aq. In this step, we were able to separate the fraction with leaf-opening activity and another one with leaf-closing activity. The fraction with leaf-closing activity was purified by HPLC using a preparative

Keywords: plants; nyctinasty; natural products; biologically active compounds.

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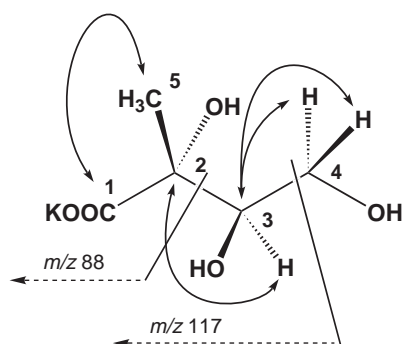


Figure 1. HMBC correlations and daughter ions observed in a linked-scan experiment of potassium 2,3,4-trihydroxy-2-methylbutanoate (**1**).

Develosil ODS-HG-5 column with 10% MeOH aq., and then Develosil ODS-HG-5 with 5% MeOH aq. to give a mixture (1.7 mg) of potassium lactate and potassium 2,3,4-trihydroxy-2-methylbutanoate (**1**) in the ratio of 1:2. We carried out further purification of **1** with HPLC with plural ODS columns, which also gave a mixture of the same compounds. Then, it was found that compound **1** was unstable around neutral or basic pH, and gradually decomposed to give lactate, which was identified by negative-mode FAB MS and ^1H NMR spectra. Then, we tried to isolate **1** by HPLC using a combination of two Develosil ODS-SR-5 columns under acidic conditions (5% MeOH aq. containing 0.5% TFA). Under this condition, we obtained **1** as a free acid form and the corresponding γ -lactone, 2,3-dihydroxy-2-methyl-4-butanolide (**2**), which was produced from **1** under acidic condition, as a mixture of 1:1. Despite many trials, further purification of **1** was unsuccessful.⁶ However, isolated potassium lactate showed no leaf-closing activity against a leaf of *L. leucocephalam*; thus, co-existing **1** was assumed to be the genuine leaf-closing substance.

We carried out a bioassay using a mixture of **1** and lactate in a ratio of 2:1 which was dissolved in a 0.1 mM phosphate buffer solution at pH 5.4. From the content of **1** in this 2:1 mixture, **1** was estimated to be effective for the leaf-closing of *L. leucocephalam* at 1×10^{-6} M. On the other hand, corresponding γ -lactone (**2**) showed no leaf-closing activity even at 1×10^{-4} M.

The structural determination of **1** was carried out by means of NMR and FAB MS experiments.⁷ HMQC and HMBC experiments gave the planar structure of **1**. A strong molecular ion corresponding to **1** was observed at m/z 189 in the positive mode FAB mass spectrum of bioactive fraction, on the other hand, m/z 149 was observed as a molecular ion in negative mode FAB MS experiment, which suggested that **1** exists as a potassium salt. Daughter ions observed in a linked-scan experiment are also shown in Fig. 1.

The relative stereochemistry of **1** was determined by using γ -lactone (**2**),⁸ which was synthesized from a mixture of **1** and **2** by treatment with 0.01 M HCl (Fig. 2). 2,3-Dihydroxy-2-methyl-4-butanolides of *syn* and *anti* relative stereochemistry were synthesized by Kobayashi et al.⁹ Thus, comparison of the ^1H NMR data of **2** with those reported by Kobayashi et al.⁹ suggested the relative stereochemistry of **2** as shown in Fig. 2. And an NOE correlation observed in **2** confirmed this result (Fig. 2). Thus, the relative stereochemistry of **1** was determined as shown in Fig. 1.

We are now examining the effect of leaf-closure with **1** on the survival of the *L. leucocephalam*. On the other hand, we have observed the leaf-opening activity in a fraction separated by the column chromatography using a TSK G3000S gel, and the isolation of the leaf-opening substance is now in progress. Either (or both) of these leaf-movement factors would be a potential herbicide of highly selective action against *L. leucocephalam*.

Acknowledgements

We are indebted to the Ministry of Education, Science, Sports and Culture (Japan) for Grant-in-Aid for Scientific Research (No. 12045259 and No. 12680598), Pioneering Research Project in Biotechnology given by the Ministry of Agriculture, Forestry and Fisheries, Kihara Foundation, Saneyoshi Foundation, Sumitomo Foundation, and Kanagawa Academy of Science and Technology Research Grant for financial support. And we thank Dr. Kazuhiko Nakamura (Biomolecules Department, National Institute of Bioscience and Human-Technology) for the collection of the plant material.

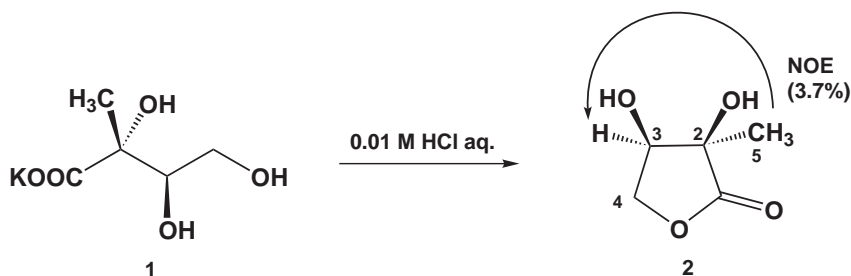


Figure 2. The preparation of γ -lactone (**2**) and the determination of stereochemistry in potassium 2,3,4-trihydroxy-2-methylbutanoate (**1**).

References

1. Darwin, C. *The Power of Movement in Plants*. Third Thousand; John Murray: London, 1882.
2. Bünning, E. *The Physiological Clock*, 3rd ed; English Univ. Press: London, 1973.
3. Ueda, M.; Yamamura, S. *Angew. Chem., Int. Ed.* **2000**, *39*, 1400–1414.
4. Ueda, M.; Sawai, Y.; Yamamura, S. *Tetrahedron* **1999**, *55*, 10925–10936.
5. Rice, E. L. *Allelopathy*, 2nd ed; Academic Press: Orlando, FL, 1984.
6. No example of the isolation of **1** has been reported. Only the detection of **1** in plant extract using GC–MS was reported in the following reference: Kringstad, R.; Singaas, A. O.; Rusten, G.; Baekkemoen, G.; Paulsen, B. S.; Nordal, A. *Phytochemistry*, **1980**, *19*, 543–545.
7. The NMR and MS experiments were carried out on a mixture of **1** and potassium lactate. Potassium 2,3,4-trihydroxy-2-methylbutanoate (**1**): ^1H NMR (400 MHz, D_2O , rt): 3.80 (1H, t, $J=5.5$ Hz, H_3), 3.57 (2H, d, $J=5.5$ Hz, H_4), 1.34 (3H, s, Me) ppm.; ^{13}C NMR (100 Mhz, D_2O , rt): 183.0 (C_1), 80.0 (C_2), 78.6 (C_3), 65.0 (C_4), 25.0 (Me) ppm.; FAB-MS (negative): $[\text{M}-\text{H}]^-$ m/z 149; FAB-MS (positive): $[\text{M}+\text{H}]^+$ m/z 189.
8. 2,3-Dihydroxy-2-methyl-4-butanolide(**2**): ^1H NMR (400 MHz, D_2O , rt): 4.58 (1H, dd, $J=3.7$, 11.0 Hz, H_{4a}), 4.29 (1H, d, $J=11.0$ Hz, H_{4b}), 4.23 (1H, d, $J=3.7$ Hz, H_3), 1.45 (3H, s, H_5) ppm.
9. Kobayashi, S.; Horibe, M.; Saito, Y. *Tetrahedron* **1994**, *50*, 9629–9642.